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**WEST**

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L15: Entry 99 of 133

File: USPT

Apr 21, 1992

DOCUMENT-IDENTIFIER: US 5106967 A

TITLE: Functional sugar substitutes with reduced calories

BSPR:

Most artificial sweeteners in use today have a greater relative sweetness than sucrose; thus, relatively small quantities are required to deliver the desired sweetness. Such low volume sweeteners may be acceptable for certain applications (e.g., beverages), however, they do not provide sufficient bulk and functionality for use in solid and semi-solid foods like baked goods and frozen desserts. In fact, even high intensity sweetener-containing beverages have a detectable reduction in their body. Two avenues have been explored to overcome this bulking problem:

BSPR:

U.S. Pat. No. 4,459,316, Bakal, issued July 10, 1984, teaches that di- and trisaccharides containing one levohexose component and at least one dextrohexose component (e.g., .alpha.-L-glucopyranosyl-D-fructofuranose) are non-caloric. These disaccharides are costly to synthesize due to the fact that they are prepared from a racemic mixture of D-hexoses and expensive L-hexoses.

BSPR:

It has now been found, that carbohydrates in the 5-C-hydroxymethyl-hexose series can be effectively used as replacements for sugar, especially in baked goods. These carbohydrate derivatives provide sucrose-like functionality (i.e., bulk, texture and stability) with significantly reduced calories compared with sucrose. In addition, many of these carbohydrate derivatives are easier to synthesize than currently available functional sugar substitutes. It is believed that they are essentially free of the significant negative physiological effects (i.e., flatus and diarrhea) generally associated with such compounds. It has also been shown that saccharides containing a 5-C-hydroxymethyl-hexose component provide similar benefits. This also holds true for the alditols of these carbohydrates (e.g., 5-C-hydroxymethyl-hexitols, 5-C-hydroxymethyl-aldohexosyl polyol derivatives, alkyl derivatives (e.g., 5-C-hydroxymethyl-aldohexosyl glycerol and 5-C-hydroxymethyl-aldohexosyl-glucitol) of the carbohydrates (i.e., alkyl 5-C-hydroxymethyl-aldohexosides), and 1,6-anhydro-.beta.-L-, and 1,6-anhydro-.beta.-D derivatives of the pyranose compounds (i.e., the bicyclic tautomeric forms) and the related derivatives of the ketohexoses.

BSPR:

The invention further encompasses food compositions (e.g., beverages, baked goods, frozen deserts and candies) containing the above-mentioned novel carbohydrates or their alditols.

BSPR:

The term "baked goods" refers to all manner of foods which are cooked (i.e., prepared using heat). These baked goods include, but are not limited to, foods prepared using dry heat (i.e., a radiant or convection oven), fried foods, boiled foods and foods heated in a microwave oven.

BSPR:

The term "food compositions" refers to and includes all manner of viand (both

sweetened and un-sweetened foods) for usage by man or animal. These food stuffs include, but are not limited to, baked goods, salted snacks, other flavored snacks, fruit drinks/mixes, frozen foods, candies, carbonated beverages, milk drinks/mixes, gelatins, puddings, fillings, breakfast cereals, breakfast bars, sauces, jams, jellies, whipped toppings, tablets, syrups, orally administered medicines, spreads, chewing gums and chocolates.

BSPR:

The term "galactose oxidase" as used herein refers to D-galactose: oxygen 6-oxidoreductase which is identified as E.C. 1.1.3.9 or as Chemical Abstracts Registry Number 9028-79-9.

BSPR:

Novel food compositions of the present invention contain from about 1% to about 99% of any of the above-mentioned compounds. Preferred embodiments of these food compositions include baked goods, fruit drinks/mixes, frozen foods, candies, carbonated beverages, milk drinks/mixes, gelatins, puddings, fillings, breakfast cereals, breakfast bars, sauces, jams, jellies, whipped toppings, tablets, syrups, orally administered medicines, spreads, chewing gums and chocolates. The most preferred food compositions are baked goods.

DEPR:

The reaction is conducted in a one liter vessel equipped with an aerator and a gentle stirrer. Sterile conditions are used to prevent enzyme deactivation by microbial contamination. The reaction is run at 4.degree. C. to minimize deactivation of galactose oxidase.

DEPR:

Methyl .beta.-D-galactopyranoside (1) is dissolved in the aerated phosphate buffer. The volume flow of air discharged by the aerator is regulated to produce an oxygen saturated solution while preventing foaming of the solution. At 4.degree. C., the galactose oxidase and catalase are added and this solution is aerated for 20 hours.

DEPR:

The reaction is conducted in a vessel equipped with a gentle stirrer and an aerator. Sterile conditions are used to prevent enzyme deactivation by microbial contamination. The reaction is run at 4.degree. C. to minimize deactivation of galactose oxidase.

DEPR:

Lactitol (23) is dissolved in the aerated phosphate buffer. At 4.degree. C., the galactose oxidase and catalase are added and this solution is aerated to maintain oxygen saturation for 20 hours.

DEPR:

The ingredients are stirred with a large spoon until well blended (about 50 strokes or 1 minute) to form a batter. The batter is poured into a lightly greased 13".times.9".times.2" pan, and then baked at 350.degree. F. for about 26.5 minutes to produce the finished brownies.

DEPR:

The ingredients are combined and the resulting dough is kneaded until uniform. Dough balls (10-13 gm) are individually placed on a lightly greased cookie tray and then baked at 350.degree. F. for 7-8 minutes to produce finished cookies.

DEPR:

The ingredients are stirred with an electric mixer to form a uniform batter. The batter is poured into a lightly greased 13".times.9".times.2" pan, and then baked at 350.degree. F. for 40 minutes to produce the finished white cake. This cake looks and tastes like a conventional white cake, but has nearly no caloric value.

DEPC:

1. Oxidation of Methyl .beta.-D-Galactopyranoside with Galactose Oxidase

DETL:

	##STR20##	Reagents	MW	Moles	Amount
		methyl .beta.-D-	194.18	0.103	20.0 g
galactopyranoside	Sigma Chemical Co.,	(No. M-6757)	Phosphate Buffer,	100 mM	--
--	412.0 ml	Catalase,	16900 units/mg	--	--
		7.5 mg	Sigma Chemical Co.,	(No.	
C-40)	<u>Galactose Oxidase</u>	--	--	9000 units	

DETL:

	##STR31##	##STR32##	Reagent	Amount
			Lactitol (23)	20.0 g (manufactured by
CCA BioChem)	Phosphate Buffer,	100 mM,	pH 7	232.0 ml
u/mg	7.00 mg	<u>Galactose Oxidase</u>	9000 units	

DETL:

	Ingredient	Amount (gms)
5-C-hydroxymethyl-.alpha.-L-arabino-hexopyranosyl-	309.8 g	D-glucitol (as prepared in Example IX)
Flour	152 g	Vegetable shortening
50 g	Cocoa	35.3 g
Starch	11.7 g	Conventional additives (flavors and a small
6.2 g	amount of <u>baking</u>	soda)
Eggs	50 g	Oil
63 g	Water	80 g

DETL:

	Ingredients	Amounts (gms)
	1,6-anhydro-5-C-hydroxymethyl-.beta.-L-	
176	altropyranose (as prepared in Example II)	Table Sugar (i.e., sucrose), 176
Flour	328	Shortening 196
Egg	96	Water 20
Conventional additives	(flavors and a small	8 amount of <u>baking</u> soda)

DETL:

	Ingredients	Amount (gms)
5-C-hydroxymethyl-.alpha.-D-xylo-hexopyrano-	133	side (as prepared in Example V)
Cake flour	107	Erythritol tetraester of olive oil
47.5	fatty acid (a polyol polyester used as a shortening)	Double-acting <u>baking</u> powder
6.7	Milk	130
Egg whites	60	Vanilla 2.5

URNM:

Bakal

**WEST**

Generate Collection

L16: Entry 1 of 5

File: USPT

May 6, 1997

DOCUMENT-IDENTIFIER: US 5626893 A

TITLE: Method of treating a divided cheese product for anticaking

## DEPR:

This example investigates the addition of dried preparations of glucose oxidase, glucose oxidase/catalase, galactose oxidase, galactose oxidase/catalase in the anticaking agent to retard growth of yeast and molds. In the prior art, glucose oxidase enzyme was combined with glucose in cellulose based anticaking agents. It was anticipated that glucose oxidase enzyme could oxidize glucose to gluconic acid and hydrogen peroxide. In so doing, it could consume a portion of the oxygen in a package of treated product. Since yeast and mold require oxygen for their growth, these additives should diminish such growth. A drawback with this procedure is that if glucose is added to the anticaking agent, and thus to the cheese, residual glucose will induce browning of pizza cheese when baked under typical pizza baking conditions. Also, simple sugars such as glucose, if they are left in cheese, encourage growth of pathogenic and unwanted bacteria. Accordingly, glucose was not added to the present anticaking agent.

**WEST**

Generate Collection

L15: Entry 103 of 133

File: USPT

Nov 12, 1991

DOCUMENT-IDENTIFIER: US 5064672 A

TITLE: Functional sugar substitutes with reduced calories

BSPR:

Most artificial sweeteners in use today have a greater relative sweetness than sucrose; thus, relatively small quantities are required to deliver the desired sweetness. Such low volume sweeteners may be acceptable for certain applications (e.g., beverages), however, they do not provide sufficient bulk and functionality for use in solid and semi-solid foods like baked goods and frozen desserts. In fact, even high intensity sweetener-containing beverages have a detectable reduction in their body. Two avenues have been explored to overcome this bulking problem:

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BSPR:

It has now been found, that carbohydrates in the 5-C-hydroxymethyl-hexose series can be effectively used as replacements for sugar, especially in baked goods. These carbohydrate derivatives provide sucrose-like functionality (i.e., bulk, texture and stability) with significantly reduced calories compared with sucrose. In addition, many of these carbohydrate derivatives are easier to synthesize than currently available functional sugar substitutes. It is believed that they are essentially free of the significant negative physiological effects (i.e., flatus and diarrhea) generally associated with such compounds. It has also been shown that saccharides containing a 5-C-hydroxymethyl-hexose component provide similar benefits. This also holds true for the alditols of these carbohydrates (e.g., 5-C-hydroxymethylhexitols, 5-C-hydroxymethyl-aldohexosyl polyol derivatives, alkyl derivatives (e.g., 5-C-hydroxymethyl-aldohexosyl glycerol and 5-C-hydroxymethyl-aldohexosyl-glucitol) of the carbohydrates (i.e., alkyl 5-C-hydroxymethyl-aldohexosides), and 1,6-anhydro-.beta.-L-, and 1,6-anhydro-.beta.-D derivatives of the pyranose compounds (i.e., the bicyclic tautomeric forms) and the related derivatives of the ketohexoses.

BSPR:

The invention further encompasses food compositions (e.g., beverages, baked goods, frozen deserts and candies) containing the above-mentioned novel carbohydrates or their alditols.

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DEPR:

The term "food compositions" refers to and includes all manner of viand (both

sweetened and un-sweetened foods) for usage by man or animal. These food stuffs include, but are not limited to, baked goods, salted snacks, other flavored snacks, fruit drinks/mixes, frozen foods, candies, carbonated beverages, milk drinks/mixes, gelatins, puddings, fillings, breakfast cereals, breakfast bars, sauces, jams, jellies, whipped toppings, tablets, syrups, orally administered medicines, spreads, chewing gums and chocolates.

DEPR:

The term "galactose oxidase" as used herein refers to D-galactose: oxygen 6-oxidoreductase which is identified as E.C. 1.1.3.9 or as Chemical Abstracts Registry Number 9028-79-9.

DEPR:

Novel food compositions of the present invention contain from about 1% to about 99% of any of the above-mentioned compounds. Preferred embodiments of these food compositions include baked goods, fruit drinks/mixes, frozen foods, candies, carbonated beverages, milk drinks/mixes, gelatins, puddings, fillings, breakfast cereals, breakfast bars, sauces, jams, jellies, whipped toppings, tablets, syrups, orally administered medicines, spreads, chewing gums and chocolates. The most preferred food compositions are baked goods.

DEPR:

The reaction is conducted in a one liter vessel equipped with an aerator and a gentle stirrer. Sterile conditions are used to prevent enzyme deactivation by microbial contamination. The reaction is run at 4.degree. C. to minimize deactivation of galactose oxidase.

DEPR:

Methyl .beta.-D-galactopyranoside (1) is dissolved in the aerated phosphate buffer. The volume flow of air discharged by the aerator is regulated to produce an oxygen saturated solution while preventing foaming of the solution. At 4.degree. C., the galactose oxidase and catalase are added and this solution is aerated for 20 hours.

DEPR:

Lactitol (23) is dissolved in the aerated phosphate buffer. At 4.degree. C., the galactose oxidase and catalase are added and this solution is aerated to maintain oxygen saturation for 20 hours.

DEPR:

The ingredients are stirred with a large spoon until well blended (about 50 strokes or 1 minute) to form a batter. The batter is poured into a lightly greased 13".times.9".times.2" pan, and then baked at 350.degree. F. for about 26.5 minutes to produce the finished brownies.

DEPR:

The ingredients are combined and the resulting dough is kneaded until uniform. Dough balls (10-13 gm) are individually placed on a lightly greased cookie tray and then baked at 350.degree. F. for 7-8 minutes to produce finished cookies.

DEPR:

The ingredients are stirred with an electric mixer to form a uniform batter. The batter is poured into a lightly greased 13".times.9".times.2" pan, and then baked at 350.degree. F. for 40 minutes to produce the finished white cake. This cake looks and tastes like a conventional white cake, by has nearly no caloric value.

DEPC:

1. Oxidation of Methyl .beta.-D-Galactopyranoside with Galactose Oxidase

DETL:

	##STR15##	Reagents	MW	Moles	Amount
		methyl .beta.-D-	194.18	0.103	20.0 g
galactopyranoside	Sigma Chemical Co.,	(No. M-6757)	Phosphate Buffer,	100 mM	--



-- 412.0 ml Catalase, 16900 units/mg -- -- 7.5 mg Sigma Chemical Co., (No. C-40) Galactose Oxidase -- -- 9000 units

## DETL:

	##STR26##	##STR27##	Reagent Amount
			Lactitol (23) 20.0 g (manufactured by
CCA BioChem) Phosphate Buffer, 100 mM, pH 7	232.0 ml	Catalase (Sigma), 16900	u/mg
7.00 mg	<u>Galactose Oxidase</u>	9000 units	

## DETL:

	Ingredient Amount (gms)
5-C-hydroxymethyl-.alpha.-L-arabino-hexopyranosyl-	309.8 g
D-glucitol (as prepared in Example IX)	Flour 152 g
Vegetable shortening	50 g
Cocoa	35.3 g
Starch	11.7 g
Conventional additives (flavors and a small	6.2 g amount of
<u>baking</u> soda)	Eggs 50 g
Oil	63 g
Water	80 g

## DETL:

	Ingredients Amounts (gms)
1,6-anhydro-5-C-hydroxymethyl-	176
.beta.-L-altropyranose (as prepared in Example II)	Table Sugar (i.e., sucrose)
176	Flour 328
Shortening	196
Egg	96
Water	20
Conventional additives (flavors and a small	8 amount of <u>baking</u> soda)

## DETL:

	Ingredients Amount (gms)
5-C-hydroxymethyl-.alpha.-D-xylo-hexopyrano-	133
side (as prepared in Example V)	Cake flour 107
Erythritol tetraester of olive oil	47.5
fatty acid (a polyol polyester used used as a shortening)	Double-acting <u>baking</u> powder 6.7
Milk	130
Egg whites	60
Vanilla	2.5

## CLPR:

10. A composition according to claim 6 where the food is selected from the group consisting of baked goods, fruit drinks/mixes, frozen food, candies, carbonated beverages, milk drinks/mixes, gelatins, puddings, fillings, breakfast cereals, breakfast bars, sauces, jams, jellies, whipped toppings, tablets, syrups, orally administered medicines, spreads, chewing gums, and chocolates.

## CLPR:

11. A composition according to claim 10 wherein the food is a baked good.

## URNM:

Bakal

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=> index bioscience

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DRUGB, DRUGLAUNCH, DRUGMONOG2, DRUGNL, ...' ENTERED AT 11:30:56 ON 22  
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=> s galactose? or lactose?

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L2 QUE GALACTOSE? OR LACTOSE?

=> s hemicellulase? or pentosanase? or xylanase? or arabinofuranosidase? or  
 mannase? or galactanase? or galactosidase?

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55 FILES HAVE ONE OR MORE ANSWERS, 56 FILES SEARCHED IN STNINDEX

L3 QUE HEMICELLULASE? OR PENTOSANASE? OR XYLANASE? OR ARABINOFURANOSIDASE?  
 OR MAMMASE? OR GALACTANASE? OR GALACTOSIDASE?

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50 FILE BIOSIS  
12\* FILE BIOTECHABS  
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15\* FILE BIOTECHNO  
11 FILE CABA  
10 FILE CANCERLIT  
64 FILE CAPLUS  
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0\* FILE CIN  
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21 FILES SEARCHED...

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0\* FILE MEDICNF  
37 FILE MEDLINE

43 FILES SEARCHED...

0\* FILE NTIS  
7 FILE SCISEARCH  
3 FILE TOXLINE  
5 FILE TOXLIT  
222 FILE USPATFULL  
6 FILE WPIDS  
6 FILE WPINDEX

26 FILES HAVE ONE OR MORE ANSWERS, 56 FILES SEARCHED IN STNINDEX

L4 QUE L3 (P) L1 (P) L2

=> s bak? or dough?

98 FILE ADISALERTS  
21 FILE ADISINSIGHT  
11713 FILE AGRICOLA  
934 FILE ANABSTR  
1041 FILE AQUASCI  
30653 FILE BIOBUSINESS  
439 FILE BIOCOMMERCE  
17873 FILE BIOSIS  
1357 FILE BIOTECHABS  
1357 FILE BIOTECHDS  
2095 FILE BIOTECHNO

11 FILES SEARCHED...

15842 FILE CABA  
1166 FILE CANCERLIT  
55834 FILE CAPLUS  
30080 FILE CEABA-VTB  
502 FILE CEN  
4182 FILE CIN  
706 FILE CONFSCI  
179 FILE CROPB  
546 FILE CROPU  
2933 FILE DDFB  
2367 FILE DDFU  
1528 FILE DGENE  
2933 FILE DRUGB  
291 FILE DRUGLAUNCH

193 FILE DRUGONOG2  
 13 FILE DRUGNL  
 2922 FILE DRUGU  
 53 FILE EMBAL  
 12557 FILE EMBASE  
 1942 FILE ESBIODBASE  
 10670 FILE FOMAD  
 4197 FILE FOREGE  
 22628 FILE FROSTI  
 34 FILES SEARCHED...  
 28227 FILE FSTA  
 18106 FILE GENBANK  
 210 FILE HEALSAFE  
 13030 FILE IFIPAT  
 7446 FILE JICST-EPLUS  
 41 FILE KOSMET  
 3330 FILE LIFESCI  
 23 FILE MEDICONF  
 19756 FILE MEDLINE  
 461 FILE NIOSHTIC  
 2334 FILE NTIS  
 316 FILE OCEAN  
 36 FILE PHAR  
 4 FILE PHIC  
 1773 FILE PHIN  
 108809 FILE PROMT  
 14313 FILE SCISEARCH  
 2737 FILE TOXLINE  
 2684 FILE TOXLIT  
 78660 FILE USPATFULL  
 54112 FILE WPIDS  
 54112 FILE WPINDEX

56 FILES HAVE ONE OR MORE ANSWERS, 56 FILES SEARCHED IN STNINDEX

L5 QUE BAK? OR DOUGH?

=> s 14 (p) 15

1 FILE BIOBUSINESS  
 0\* FILE BIOCOMMERCE  
 1 FILE BIOSIS  
 0\* FILE BIOTECHABS  
 0\* FILE BIOTECHDS  
 0\* FILE BIOTECHNO  
 0\* FILE CEABA-VTB  
 0\* FILE CIN  
 20 FILES SEARCHED...  
 0\* FILE ESBIODBASE  
 0\* FILE FOMAD  
 0\* FILE FOREGE  
 3\* FILE FROSTI  
 0\* FILE FSTA  
 0\* FILE KOSMET  
 41 FILES SEARCHED...  
 0\* FILE MEDICONF  
 0\* FILE NTIS

3 FILES HAVE ONE OR MORE ANSWERS, 56 FILES SEARCHED IN STNINDEX

L6 QUE L4 (P) L5

=> d rank

F1 3\* FROSTI  
 F2 1 BIOBUSINESS

=> file f1-f3

COST IN U.S. DOLLARS

SINCE FILE

TOTAL

ENTRY

SESSION

FULL ESTIMATED COST

5.40

5.55

FILE 'FROSTI' ENTERED AT 11:37:54 ON 22 JAN 2001

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FILE 'BIOBUSINESS' ENTERED AT 11:37:54 ON 22 JAN 2001

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FILE 'BIOSIS' ENTERED AT 11:37:54 ON 22 JAN 2001

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=> s 16

PROXIMITY OPERATOR LEVEL NOT CONSISTENT WITH  
FIELD CODE - 'AND' OPERATOR ASSUMED 'L3 (P) L1'  
PROXIMITY OPERATOR LEVEL NOT CONSISTENT WITH  
FIELD CODE - 'AND' OPERATOR ASSUMED 'L1 (P) L2'  
PROXIMITY OPERATOR LEVEL NOT CONSISTENT WITH  
FIELD CODE - 'AND' OPERATOR ASSUMED 'L4 (P) L5'  
L7 5 L6

=> dup rem 17

PROCESSING COMPLETED FOR L7

L8 4 DUP REM L7 (1 DUPLICATE REMOVED)

=> d 1-4 ab,bib

L8 ANSWER 1 OF 4 FROSTI COPYRIGHT 2001 LFRA

AB Arabinogalactan-peptide (AGP) is a group of water-soluble macromolecules with a highly branched structure. The amino acid composition of the peptidic fraction could provide functional properties through serving as a link to the carbohydrate fraction. The possible use of wheat flour AGP or its degradation products as a substrate for an oxidative enzyme was evaluated with **galactose oxidase**. This enzyme could be an alternative oxidative enzyme for use in bread-making. The composition and depolymerization of wheat flour AGP were determined. The effects of selected enzymic activities on oxidation were also evaluated. A crude liquid enzyme preparation from *Aspergillus niger* displayed activities capable of depolymerizing wheat flour AGP to galactobiose, **galactose** and arabinose. It could also produce substrate from the wheat flour AGP, associated with alpha-L-**arabinofuranosidase**.

AN 496464 FROSTI

TI Production of substrate for **galactose oxidase** by depolymerization of an arabinogalactan-peptide from wheat flour.

AU Schroder M.; Soe J.B.; Zargahi M.R.; Rouau X.

SO Journal of Agricultural and Food Chemistry, 1999, (April), 47 (4), 1483-1488 (19 ref.)

ISSN: 0021-8561

DT Journal

LA English

SL English

L8 ANSWER 2 OF 4 BIOBUSINESS COPYRIGHT 2001 BIOSIS DUPLICATE 1

AN 96:71597 BIOBUSINESS

DN 0836458

TI Application of oxidoreductases in baking: Impact of some oxidoreductases on gluten structure and dough rheology.



AU Somers W A C; Or R; Van Der Lugt J P  
CS TNO Nutrition Food Res. Inst., P.O. Box 360, 3700 Zeist, Netherlands  
SO Cereal Foods World, (1996) Vol.41, No.7, P.550.  
81st Annual Meeting of the American Association of Cereal Chemistry,  
Baltimore, Maryland, USA, September 15-19, 1996. CEREAL FOODS WORLD.

ISSN:

0146-6283.

DT CONFERENCE

FS NONUNIQUE

LA ENGLISH

L8 ANSWER 3 OF 4 FROSTI COPYRIGHT 2001 LFRA

AB A composition for **dough** and bread improvement is disclosed. It incorporates an enzyme with **galactose oxidase** activity and an oxidizable substrate for this enzyme. It is said to improve the rheological properties of flour **doughs** and the quality characteristics of bread products. Desirable quality characteristics include soft crumb structure, high specific volume, and freedom from staling within the expected shelf-life of fresh bread.

**Galactose oxidase** acts as an oxidoreductase, and its use overcomes problems associated with use of cellulases or **hemicellulases** in flour **doughs**. Because the natural **galactose** content of cereal flours is very low, it is beneficial to include an oxidizable substrate in the formulation.

AN 489211 FROSTI

TI A composition comprising an enzyme having **galactose oxidase** activity and use thereof.

IN Rouau X.; Schroder M.; Soe J.B.

PA Danisco A/S

SO PCT Patent Application

PI WO 9903351 A1

AI 19980716

PRAI Denmark 19970718

United States 19970722

DT Patent

LA English

SL English

L8 ANSWER 4 OF 4 FROSTI COPYRIGHT 2001 LFRA

AB A composition for **dough** and bread improvement is disclosed. It incorporates an enzyme with **galactose oxidase** activity and an oxidizable substrate for this enzyme. It is said to improve the rheological properties of flour **doughs** and the quality characteristics of bread products. Desirable quality characteristics include soft crumb structure, high specific volume, and freedom from staling within the expected shelf-life of fresh bread.

**Galactose oxidase** acts as an oxidoreductase, and its use overcomes problems associated with use of cellulases or **hemicellulases** in flour **doughs**. Because the natural **galactose** content of cereal flours is very low, it is beneficial to include an oxidizable substrate in the formulation.

AN 526332 FROSTI

TI A composition comprising an enzyme having **galactose oxidase** activity and use thereof.

IN Rouau X.; Schroder M.; Soe J.B.

PA Danisco A/S

SO European Patent Application

PI EP 999752 A1

WO 9903351 19990128

AI 19980716

PRAI Denmark 19970718

United States 19970722

DT Patent

LA English

SL English

=> in dex bioscience

IN IS NOT A RECOGNIZED COMMAND

The previous command name entered was not recognized by the system.  
For a list of commands available to you in the current file, enter  
"HELP COMMANDS" at an arrow prompt (=>).

=> index bioscience

FILE 'DRUGMONOG' ACCESS NOT AUTHORIZED  
COST IN U.S. DOLLARS

SINCE FILE	TOTAL
ENTRY	SESSION
6.93	12.48

FULL ESTIMATED COST

INDEX 'ADISALERTS, ADISINSIGHT, AGRICOLA, ANABSTR, AQUASCI, BIOBUSINESS,  
BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO, CABA, CANCERLIT,  
CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPU, DDFB, DDFU, DGENE,  
DRUGB, DRUGLAUNCH, DRUGMONOG2, DRUGNL, ...' ENTERED AT 11:39:07 ON 22  
JAN 2001

56 FILES IN THE FILE LIST IN STNINDEX

Enter SET DETAIL ON to see search term postings or to view  
search error messages that display as 0\* with SET DETAIL OFF.

=> s 14 and 15

1	FILE BIOBUSINESS
0*	FILE BIOCOMMERCE
1	FILE BIOSIS
0*	FILE BIOTECHABS
0*	FILE BIOTECHDS
0*	FILE BIOTECHNO
14 FILES SEARCHED...	
0*	FILE CEABA-VTB
0*	FILE CIN
0*	FILE ESBIOWASE
0*	FILE FOMAD
0*	FILE FOREGE
3*	FILE FROSTI
0*	FILE FSTA
35 FILES SEARCHED...	
0*	FILE KOSMET
0*	FILE MEDICONE
0*	FILE NTIS
66	FILE USPATFULL

4 FILES HAVE ONE OR MORE ANSWERS, 56 FILES SEARCHED IN STNINDEX

L9 QUE L4 AND L5

=> d rank

F1	66	USPATFULL
F2	3*	FROSTI
F3	1	BIOBUSINESS
F4	1	BIOSIS

=> s 11 (p) ((12 or 13))

56	FILE AGRICOLA
41	FILE ANABSTR
2	FILE AQUASCI
16	FILE BIOBUSINESS

3\* FILE BIOCOMMERCE  
 1028 FILE BIOSIS  
 110\* FILE BIOTECHABS  
 110\* FILE BIOTECHDS  
 242\* FILE BIOTECHNO  
 91 FILE CABA  
 293 FILE CANCERLIT  
 1305 FILE CAPLUS  
 22\* FILE CEABA-VTB  
 1 FILE CEN  
 1\* FILE CIN  
 37 FILE CONFSCI  
 21 FILE DDFB  
 6 FILE DDFU  
 13 FILE DGENE  
 23 FILES SEARCHED...  
 21 FILE DRUGB  
 16 FILE DRUGU  
 1 FILE EMBAL  
 688 FILE EMBASE  
 88\* FILE ESBIODBASE  
 0\* FILE FOMAD  
 0\* FILE FOREGE  
 20\* FILE FROSTI  
 23\* FILE FSTA  
 7 FILE GENBANK  
 1 FILE HEALSAFE  
 100 FILE IFIPAT  
 135 FILE JICST-EPLUS  
 2\* FILE KOSMET  
 206 FILE LIFESCI  
 0\* FILE MEDICONF  
 833 FILE MEDLINE  
 4 FILE NIOSHTIC  
 5\* FILE NTIS  
 2 FILE PHIN  
 6 FILE PROMT  
 548 FILE SCISEARCH  
 51 FILES SEARCHED...  
 60 FILE TOXLINE  
 72 FILE TOXLIT  
 691 FILE USPATFULL  
 88 FILE WPIDS  
 88 FILE WPINDEX

43 FILES HAVE ONE OR MORE ANSWERS, 56 FILES SEARCHED IN STNINDEX

L10 QUE L1 (P) ((L2 OR L3))

=> s 110 and 15

1 FILE BIOBUSINESS  
 0\* FILE BIOCOMMERCE  
 1 FILE BIOSIS  
 0\* FILE BIOTECHABS  
 0\* FILE BIOTECHDS  
 0\* FILE BIOTECHNO  
 3 FILE CAPLUS  
 1\* FILE CEABA-VTB  
 0\* FILE CIN  
 22 FILES SEARCHED...  
 0\* FILE ESBIODBASE  
 0\* FILE FOMAD  
 0\* FILE FOREGE  
 3\* FILE FROSTI  
 0\* FILE FSTA

1 FILE GENBANK  
2 FILE IFIPAT  
0\* FILE KOSMET  
0\* FILE MEDICONF  
0\* FILE NTIS  
48 FILES SEARCHED...  
129 FILE USPATFULL  
2 FILE WPIDS  
2 FILE WPINDEX

10 FILES HAVE ONE OR MORE ANSWERS, 56 FILES SEARCHED IN STNINDEX

L11 QUE L10 AND L5

=> d rank

F1	129	USPATFULL
F2	3	CAPLUS
F3	3*	FROSTI
F4	2	IFIPAT
F5	2	WPIDS
F6	2	WPINDEX
F7	1	BIOBUSINESS
F8	1	BIOSIS
F9	1	GENBANK
F10	1*	CEABA-VTB

=> file f2-f10

COST IN U.S. DOLLARS	SINCE FILE ENTRY	TOTAL SESSION
FULL ESTIMATED COST	3.15	15.63

FILE 'CAPLUS' ENTERED AT 11:43:01 ON 22 JAN 2001  
USE IS SUBJECT TO THE TERMS OF YOUR STN CUSTOMER AGREEMENT.  
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FILE 'FROSTI' ENTERED AT 11:43:01 ON 22 JAN 2001  
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COPYRIGHT (C) 2001 IFI CLAIMS(R) Patent Services (IFI)

FILE 'WPIDS' ENTERED AT 11:43:01 ON 22 JAN 2001  
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FILE 'WPINDEX' ACCESS NOT AUTHORIZED

FILE 'BIOBUSINESS' ENTERED AT 11:43:01 ON 22 JAN 2001  
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FILE 'BIOSIS' ENTERED AT 11:43:01 ON 22 JAN 2001  
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FILE 'GENBANK' ENTERED AT 11:43:01 ON 22 JAN 2001

FILE 'CEABA-VTB' ENTERED AT 11:43:01 ON 22 JAN 2001  
COPYRIGHT (c) 2001 DECHEMA eV

=> s 111

PROXIMITY OPERATOR LEVEL NOT CONSISTENT WITH  
FIELD CODE - 'AND' OPERATOR ASSUMED 'L1 (P) '  
PROXIMITY OPERATOR LEVEL NOT CONSISTENT WITH

FIELD CODE - 'AND' OPERATOR ASSUMED 'L1 (P) '  
L12 14 L11

=> dup rem l12

DUPLICATE IS NOT AVAILABLE IN 'GENBANK'.  
ANSWERS FROM THESE FILES WILL BE CONSIDERED UNIQUE  
PROCESSING COMPLETED FOR L12  
L13 13 DUP REM L12 (1 DUPLICATE REMOVED)

=> d 1-13 ab,bib

NO VALID FORMATS ENTERED FOR FILE 'GENBANK'

In a multifile environment, each file must have at least one valid  
format requested. Refer to file specific help messages or the  
STNGUIDE file for information on formats available in individual  
files.

REENTER DISPLAY FORMAT FOR ALL FILES (FILEDEFAULT):all

L13 ANSWER 1 OF 13 WPIDS COPYRIGHT 2001 DERWENT INFORMATION LTD

AN 1999-131751 [11] WPIDS

DNC C1999-038439

TI A **dough** and bread improving composition comprising a  
**galactose oxidase** and a substrate for it - useful for  
improving the rheological characteristics of flour **dough** with a  
**dough** strengthening effect, without stickiness and/or slackness.

DC D11 D16

IN ROUAU, X; SCHRODER, M; SOE, J B

PA (DANI-N) DANISCO AS

CYC 83

PI WO 9903351 A1 19990128 (199911)\* EN 41p A21D008-04

RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW NL

OA PT SD SE SZ UG ZW

W: AL AM AT AU AZ BA BB BG BR BY CA CH CN CU CZ DE DK EE ES FI GB GE

GH GM HR HU ID IL IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MD MG

MK MN MW MX NO NZ PL PT RO RU SD SE SG SI SK SL TJ TM TR TT UA UG

US UZ VN YU ZW

AU 9883347 A 19990210 (199925) A21D008-04

EP 999752 A1 20000517 (200028) EN A21D008-04

R: AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE

ADT WO 9903351 A1 WO 1998-DK335 19980716; AU 9883347 A AU 1998-83347  
19980716;

EP 999752 A1 EP 1998-933577 19980716, WO 1998-DK335 19980716

FDT AU 9883347 A Based on WO 9903351; EP 999752 A1 Based on WO 9903351

PRAI US 1997-53451 19970722; DK 1997-878 19970718

IC ICM A21D008-04

ICS A23L001-16

AB WO 9903351 A UPAB: 19990316

A **dough** and bread improving composition comprises (a) an enzyme  
having **galactose oxidase** activity, and (b) an  
oxidisable substrate for (a) and/or an enzyme which can convert a  
compound  
into this substrate. Also claimed is a method of preparing a flour  
**dough**.

USE - The composition is useful for improving the rheological  
characteristics of flour **dough** with a **dough**  
strengthening effect, without stickiness and/or slackness

ADVANTAGE - Any type of flour **dough** can be used, e.g. wheat  
flour based bread products, noodle products, alimentary paste product,  
etc.

Dwg.0/4

FS CPI

FA AB

MC CPI: D01-B01; D01-B02A; D05-A02A

L13 ANSWER 2 OF 13 FROSTI COPYRIGHT 2001 LFRA

AN 496464 FROSTI  
TI Production of substrate for **galactose oxidase** by  
depolymerization of an arabinogalactan-peptide from wheat flour.  
AU Schroder M.; Soe J.B.; Zargahi M.R.; Rouau X.  
SO Journal of Agricultural and Food Chemistry, 1999, (April), 47 (4),  
1483-1488 (19 ref.)  
ISSN: 0021-8561

DT Journal

LA English

SL English

AB Arabinogalactan-peptide (AGP) is a group of water-soluble macromolecules with a highly branched structure. The amino acid composition of the peptidic fraction could provide functional properties through serving as a link to the carbohydrate fraction. The possible use of wheat flour AGP or its degradation products as a substrate for an oxidative enzyme was evaluated with **galactose oxidase**. This enzyme could be an alternative oxidative enzyme for use in bread-making. The composition and depolymerization of wheat flour AGP were determined. The effects of selected enzymic activities on oxidation were also evaluated. A crude liquid enzyme preparation from *Aspergillus niger* displayed activities capable of depolymerizing wheat flour AGP to galactobiose, **galactose** and arabinose. It could also produce substrate from the wheat flour AGP, associated with alpha-L-arabinofuranosidase.

SH CEREAL PRODUCTS

CT ARABINO GALACTAN PEPTIDE; ASPERGILLUS; ASPERGILLUS NIGER; **BAKERY**  
PRODUCTS; BREAD; CEREAL FLOURS; CEREAL PRODUCTS; COMPOSITION;  
DEPOLYMERIZATION; ENZYMIC ACTIVITY; FLOURS; FUNGI; **GALACTOSE**  
**OXIDASE**; MICROORGANISMS; SUBSTRATES; WHEAT FLOUR; WHEAT PRODUCTS

DED 17 Jun 1999

L13 ANSWER 3 OF 13 CAPLUS COPYRIGHT 2001 ACS

AN 1998:454862 CAPLUS

DN 129:215945

TI Application of oxidoreductases in **baking**: impact on gluten structure and **dough** rheology

AU Van Der Lugt, J. P.; Somers, W. A. C.; Lichtendonk, W.; Orsel, R.

CS TNO Nutrition and Food Research Institute, Zeist, 3700 AJ, Neth.

SO Eur. Symp. Enzymes Grain Process., Proc., 1st (1997), Meeting Date 1996,  
164-176. Editor(s): Angelino, S. A. G. F. Publisher: TNO Nutrition and  
Food Research Institute, Zeist, Neth.

CODEN: 66KVAR

DT Conference

LA English

CC 17-11 (Food and Feed Chemistry)

AB Rheol. measurements of **dough** and glutenin macro polymer systems were used to study effects of enzymes. Glucose oxidase improved the complex modulus ( $G^*$ ). **Galactose oxidase** under favorable conditions resulted in better **dough** rigidity and increased the elastic behavior of the **dough**. Lignin peroxidase gave the opposite effect. Lipoxxygenase increased  $G^*$ , presumably due to oxidn. of protein polymers.

ST gluten rheol **dough** enzyme; glucose oxidase gluten rheol  
**dough**; **galactose oxidase** gluten rheol  
**dough**; lignin peroxidase gluten rheol **dough**;  
lipoxxygenase gluten rheol **dough**

IT **Baking**

**Dough**

Food elasticity

Food rheology

(oxidoreductases in relation to gluten structure and **dough** rheol.)

IT Glutens

RL: BPR (Biological process); PRP (Properties); BIOL (Biological study);

PROC (Process)

(oxidoreductases in relation to gluten structure and **dough**)

rheol.)  
IT 9001-37-0, Glucose oxidase 9028-79-9, Galactose  
oxidase 9029-60-1, Lipoxxygenase 42613-30-9, Lignin peroxidase  
RL: BAC (Biological activity or effector, except adverse); FFD (Food or  
feed use); BIOL (Biological study); USES (Uses)  
(oxidoreductases in relation to gluten structure and dough  
rheol.)

L13 ANSWER 4 OF 13 CAPLUS COPYRIGHT 2001 ACS

AN 1996:365938 CAPLUS

DN 125:32391

TI Anticaking agent for dairy products

IN Reddy, Malireddy S.

PA USA

SO PCT Int. Appl., 56 pp.

CODEN: PIXXD2

DT Patent

LA English

IC ICM A23C019-14

CC 17-8 (Food and Feed Chemistry)

FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 9611581	A1	19960425	WO 1995-US12860	19951017
	W: AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LT, LU, LV, MD, MG, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TT				
	RW: KE, MW, SD, SZ, UG, AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG				
	US 5626893	A	19970506	US 1994-324897	19941018
	AU 9538278	A1	19960506	AU 1995-38278	19951017
	AU 689335	B2	19980326		
	EP 786941	A1	19970806	EP 1995-936266	19951017
	R: DE, DK, FR, GB				

PRAI US 1994-324897 19941018

WO 1995-US12860 19951017

AB An anticaking agent which reduces the stickiness of the chunked, diced,  
or

shredded cheese and improves the functionality of cheese is formulated of  
fine mesh vegetable flour, bentonite, cellulose, and antimycotic agents

or  
bacterial cultures. This anticaking agent also will reduce the yeast and  
mold growth. This discovery is also extended to include various flavors,  
colors, enzymes and other supplements into the anticaking agent, which

are  
to be ultimately added to the cheese.

ST dairy product anticaking agent

IT Agglomeration preventers

Bifidobacterium bifidum

Capsicum

Capsicum frutescens

Cheese

Corn

Dill

Emulsifying agents

Flavor

Flours and Meals

Fungicides and Fungistats

Garlic

Lactobacillus acidophilus

Lactobacillus bulgaricus

Lactobacillus helveticus

Lactobacillus lactis

Lactococcus lactis

Leuconostoc mesenteroides  
 Milk  
 Oregano  
 Pediococcus acidilactici  
 Pediococcus cerevisiae  
 Pediococcus pentosaceus  
 Potato  
 Propionibacterium freudenreichii  
 Propionibacterium shermanii  
 Rice  
 Streptococcus salivarius  
 Wheat  
 Whey  
 Yeast  
 (anticaking agent for dairy products)

IT Vitamins  
 RL: BAC (Biological activity or effector, except adverse); FFD (Food or feed use); BIOL (Biological study); USES (Uses)  
 (anticaking agent for dairy products)

IT Bentonite, biological studies  
 RL: FFD (Food or feed use); PEP (Physical, engineering or chemical process); BIOL (Biological study); PROC (Process); USES (Uses)  
 (anticaking agent for dairy products)

IT Carboxylic acids, biological studies  
 RL: FFD (Food or feed use); PEP (Physical, engineering or chemical process); BIOL (Biological study); PROC (Process); USES (Uses)  
 (anticaking agent for dairy products)

IT Phosphates, biological studies  
 RL: FFD (Food or feed use); PEP (Physical, engineering or chemical process); BIOL (Biological study); PROC (Process); USES (Uses)  
 (anticaking agent for dairy products)

IT Flavoring materials  
 (spice; anticaking agent for dairy products)

IT Flavoring materials  
 (Cheddar cheese; anticaking agent for dairy products)

IT Cheese  
 (Monterey Jack; anticaking agent for dairy products)

IT Cheese  
 (Mozzarella; anticaking agent for dairy products)

IT Cheese  
 (Parmesan; anticaking agent for dairy products)

IT Cheese  
 (Romano; anticaking agent for dairy products)

IT Flavoring materials  
 (cheese; anticaking agent for dairy products)

IT Food functional properties  
 (emulsifying; anticaking agent for dairy products)

IT Food functional properties  
 (foaming; anticaking agent for dairy products)

IT Flavoring materials  
 (fruit; anticaking agent for dairy products)

IT **Bakery products**  
 (pizza; anticaking agent for dairy products)

IT 64-19-7, Acetic acid, biological studies 65-85-0, Benzoic acid, biological studies 79-09-4D, Propionic acid, salts 110-44-1D, Sorbic acid, salts 137-40-6, Sodium propionate 431-03-8, Diacetyl 1406-16-2, Vitamin D 7681-93-8, Natamycin 9001-05-2, Catalase 9001-37-0, Glucose oxidase 9028-79-9, **Galactose oxidase** 9031-11-2, Lactase 11103-57-4, Vitamin A  
 RL: BAC (Biological activity or effector, except adverse); FFD (Food or feed use); BIOL (Biological study); USES (Uses)  
 (anticaking agent for dairy products)

IT 50-99-7, Dextrose, biological studies  
 RL: FFD (Food or feed use); BIOL (Biological study); USES (Uses)  
 (anticaking agent for dairy products)

IT 50-21-5, Lactic acid, biological studies 50-81-7, Ascorbic acid,



biological studies 68-04-2, Sodium citrate 77-9, Citric acid,  
biological studies 471-34-1, Calcium carbonate, biological studies  
1393-63-1, Annatto 7601-54-9, Sodium phosphate 7664-38-2, Phosphoric  
acid, biological studies 9004-34-6, Cellulose, biological studies  
9005-25-8, Starch, biological studies 9016-00-6, Polydimethyl siloxane  
9050-36-6, Maltodextrin 13478-98-3, Hexametaphosphate  
RL: FFD (Food or feed use); PEP (Physical, engineering or chemical  
process); BIOL (Biological study); PROC (Process); USES (Uses)  
(anticaking agent for dairy products)

IT 1318-93-0, Sodium montmorillonite, biological studies  
RL: FFD (Food or feed use); PEP (Physical, engineering or chemical  
process); BIOL (Biological study); PROC (Process); USES (Uses)  
(sodium-exchanged; anticaking agent for dairy products)

L13 ANSWER 5 OF 13 BIOBUSINESS COPYRIGHT 2001 BIOSIS DUPLICATE 1

AN 96:71597 BIOBUSINESS

DN 0836458

TI Application of oxidoreductases in **baking**: Impact of some  
oxidoreductases on gluten structure and **dough** rheology.

AU Somers W A C; Orsel R; Van Der Lugt J P

CS TNO Nutrition Food Res. Inst., P.O. Box 360, 3700 AJ Zeist, Netherlands

SO Cereal Foods World, (1996) Vol.41, No.7, P.550.

81st Annual Meeting of the American Association of Cereal Chemistry,  
Baltimore, Maryland, USA, September 15-19, 1996. CEREAL FOODS WORLD.

ISSN:

0146-6283.

DT CONFERENCE

FS NONUNIQUE

LA ENGLISH

CC 40200 **BAKING** TECHNOLOGY; 40300 CEREAL CHEMISTRY; 40400  
CHEMICAL & PHYSICAL PROPERTIES OF FOODS; 40900 FOOD PREPARATION,  
PROCESSING & STORAGE

ST MEETING ABSTRACT; HYDROGEN PEROXIDE; **GALACTOSE OXIDASE**  
; GLUCOSE OXIDASE; **HEMICELLULASE**; POLYMER SIZE; DISULFIDE BOND;  
LOAF VOLUME; **BAKERY** PRODUCTS; GRAIN PRODUCTS; FOOD PROCESSING;  
FOOD CHEMISTRY

RN 7722-84-1 (HYDROGEN PEROXIDE)

9001-37-0 (GLUCOSE OXIDASE)

9025-56-3 (**HEMICELLULASE**)

9028-79-9 (**GALACTOSE OXIDASE**)

16734-12-6 (DISULFIDE)

L13 ANSWER 6 OF 13 CAPLUS COPYRIGHT 2001 ACS

AN 1994:279904 CAPLUS

DN 120:279904

TI Stabilized chewable antimicrobial foodstuff for animal

IN Montgomery, Robert E.

PA USA

SO PCT Int. Appl., 30 pp.

CODEN: PIXXD2

DT Patent

LA English

IC ICM A61K007-28

ICS A61K037-50

CC 62-7 (Essential Oils and Cosmetics)

Section cross-reference(s): 18

FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 9405252	A1	19940317	WO 1993-US8086	19930827
W: AU, CA				
RW: AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
US 5310541	A	19940510	US 1992-936929	19920827
EP 658096	A1	19950621	EP 1993-921221	19930827
EP 658096	B1	19991103		
R: DE, ES, FR, GB, IT, NL				

ES 2141171 3 20000316 ES 1993-921 19930827  
PRAI US 1992-936929 19920827  
WO 1993-US8086 19930827  
AB The invention is an animal chew which contains one or more enzymes and substrates for the purpose of generating antimicrobial compds. upon contact with an animal's saliva. The animal chew, made of rawhide, biscuit or dried animal food is provided with an oxidoreductase enzyme and substrate, such as glucose oxidase and glucose, which produces H2O2 upon being chewed. A catalase may be provided to stabilize the system and prevent premature activation of the enzyme/substrate system. A peroxidase and halide or pseudohalide ion combination may be provided to enhance the antimicrobial effect of the invention.  
ST chewable antimicrobial animal dentifrice enzyme peroxide  
IT Chlorides, biological studies  
Halides  
Iodides, biological studies  
Pseudohalides  
RL: BIOL (Biological study)  
(antimicrobial animal chewing foodstuff contg. oxidoreductase and peroxidase and, for inhibiting oral pathogens)  
IT Dentifrices  
(bactericidal, chewable, for animal, oxidoreductase and enzyme substrate in)  
IT Hide substances  
(raw-, antimicrobial animal chewing foodstuff contg. oxidoreductase and peroxidase and enzyme substrates and, for inhibiting oral pathogens)  
IT Bakery products  
(biscuits, for animal, antimicrobial animal chewing foodstuff contg. oxidoreductase and peroxidase and enzyme substrates and, for inhibiting oral pathogens)  
IT Food  
(dry, for animal, antimicrobial animal chewing foodstuff contg. oxidoreductase and peroxidase and enzyme substrates and, for inhibiting oral pathogens)  
IT 9003-99-0, Lactoperoxidase 9055-20-3, Chloroperoxidase  
RL: BIOL (Biological study)  
(antimicrobial animal chewing foodstuff contg. oxidoreductase and enzyme substrates and, for inhibiting oral pathogens)  
IT 333-20-0, Potassium thiocyanate 540-72-7, Sodium thiocyanate 1762-95-4, Ammonium thiocyanate 7647-14-5, Sodium chloride, biological studies 7681-11-0, Potassium iodide, biological studies  
RL: BIOL (Biological study)  
(antimicrobial animal chewing foodstuff contg. oxidoreductase and peroxidase and, for inhibiting oral pathogens)  
IT 50-99-7, D-Glucose, biological studies 51-67-2, Tyramine 59-23-4, d-Galactose, biological studies 64-17-5, Ethanol, biological studies 69-89-6, Xanthine 75-07-0, Acetaldehyde, biological studies 79-14-1, Glycolic acid, biological studies 79-33-4, L-Lactic acid, biological studies 87-79-6, L-Sorbose 95-55-6, 2-Aminophenol 110-60-1, 1,4-Diaminobutane 123-72-8, Butyraldehyde 154-17-6, 2-Deoxy-D-glucose 1783-96-6, D-Aspartic acid 6893-26-1, D-Glutamic acid 10516-09-3 13748-90-8, L-2-Hydroxyisocaproic acid 14474-04-5 22956-40-7 32746-79-5 106623-56-7  
RL: BIOL (Biological study)  
(antimicrobial animal chewing foodstuff contg. oxidoreductase and, for inhibiting oral pathogens)  
IT 9000-88-8, D-Amino acid oxidase 9000-89-9, L-Amino acid oxidase 9001-37-0, Glucose oxidase 9001-53-0, Diamine oxidase 9001-66-5, Monoamine oxidase 9028-71-1, Glycollate oxidase 9028-72-2, Lactate oxidase 9028-78-8 9028-79-9, Galactose oxidase 9029-21-4, Pyridoxaminephosphate oxidase 9029-38-3, Sulfite oxidase

RL: BIOL (Biological study)  
(antimicrobial animal chewing foodstuff contg. for inhibiting oral  
pathogens)

L13 ANSWER 7 OF 13 CEABA-VTB COPYRIGHT 2001 DECHEMA  
AN 1995(07):1196 CEABA-VTB FS B  
DN CEABA: 1995:5081821  
TI Microbial cometabolism of sucralose, a chlorinated disaccharide, in  
environmental samples  
AU Labare, M.P.; Alexander, M. (Cornell Univ., Ithaca NY, USA)  
SO Appl. Microbiol. Biotechnol. (1994) 42(1), p.173-178, 5f,1t,12l  
CODEN: AMBIDG ISSN: 0175-7598  
DT Journal  
LA English  
AB The paper reports on investigations into the mineralization of sucralose  
(4-chloro-4-deoxy-.alpha.,D-fructofuranoside). During rapid  
mineralization in soil and slow mineralization in lake water, a  
corresponding unsaturated aldehyde appeared to be the metabolic product.  
Organic products from the disaccharide were not detected in sewage under  
aerobic conditions and little or no CO2 was detected under anaerobic  
conditions. Sucralose carbon was not found in the cells of  
sucralose-metabolizing bacteria or the microbial mass of sewage. A  
**galactose oxidase** slowly metabolized the chlorinated  
disaccharide and it is concluded that sucralose transformation is the  
result of microbial cometabolism. (Hryniewicz)  
CC 9442 Water treatment  
141 Organic chemistry  
145 Microbial and biochemical reactions  
9141 Bacteria, cyanobacterial (Prokaryota)  
CT BACTERIA; ENVIRONMENTAL POLLUTION; ENZYME; TRANSFORMATION

L13 ANSWER 8 OF 13 IFIPAT COPYRIGHT 2001 IFI  
AN 2417486 IFIPAT;IFIUDB;IFICDB  
TI STABILIZED ENZYMATIC ANTIMICROBIAL COMPOSITIONS; ADDITION OF CATALASE  
TOGETHER WITH OXIDOREDUCTASE PREVENTS FORMATION OF HYDROGEN PEROXIDE SO  
PREVENT THE SALIVARY PEROXIDASE SYSTEM PRODUCING HYPOTHIOCYANITE  
INF Montgomery, Robert E, 8916 Hollywood Hills Rd, Los Angeles, CA, 90046  
IN Montgomery Robert E  
PAF Unassigned  
PA Unassigned Or Assigned To Individual (68000)  
EXNAM Rose, Shep K

AG Blakely, Sokoloff, Taylor, Zafman  
PI US 5262151 19931116 (CITED IN 002 LATER PATENTS)  
AI US 1992-934772 19920824  
XPD 25 Nov 2011  
RLI US 1991-797776 19911125 CONTINUATION-IN-PART 5176899  
FI US 5262151 19931116  
US 5176899

DT UTILITY  
FS CHEMICAL  
AB A stabilized aqueous composition capable of producing or, in the  
presence  
of saliva or other humoral fluid, leading to the production of  
antimicrobially effective concentrations of hypothyocyanite ions (OSCN-)  
are herein described. The composition contains an oxidoreductase enzyme  
and its specific substrate, for the purpose of producing hydrogen  
peroxide of at least the minimum effective concentration, and in  
addition, catalase for the destruction of hydrogen peroxide to prevent  
premature oxidoreductase enzyme decomposition. Optionally, a peroxidase  
enzyme may be included to act upon the aforementioned hydrogen peroxide,  
thereby oxidizing thiocyanate ions to produce the antimicrobial  
concentrations of hypothyocyanite ions (OSCN).

CLMN 23  
ECLM 1. An antimicrobial dentifrice composition made by the process  
comprising  
the steps of: providing a fluid carrier comprising an oxidoreductase

enzyme, an oxidoreductase enzyme substrate and a catalase, wherein said enzyme and substrate form hydrogen peroxide when reacted together in the presence of oxygen, said hydrogen peroxide being formed at a rate of at least 100 micromoles per liter per minute, said oxidoreductase enzyme being present in said composition in the amount of at least 1.0 Titrimetric Unit per gram of dentifrice, and wherein said catalase is provided in a ratio of about 50 Titrimetric Units oxidoreductase enzyme to 1.0 Baker Unit of catalase, to 1.0 Titrimetric Units oxidoreductase enzyme to 1.0 Baker Unit of catalase to substantially minimize the amount of hydrogen peroxide produced in said composition during storage; and storing said mixture in an oxygen impervious container.

ACLM 2. The composition of claim 1 wherein catalase is provided in an amount in the ratio range of 50 Titrimetric Units of oxidoreductase to 1.0

Baker Unit of catalase, to 1.0 Titrimetric Units of oxidoreductase to 1.0 Baker Unit of catalase.

3. The composition of claim 2 wherein the ratio of oxidoreductase to catalase is about 6.0 Titrimetric Units of oxidoreductase to 1.0

Baker Unit of catalase.

4. The composition of claim 1 further comprising a peroxidase enzyme for oxidizing thiocyanate ions to hypothiocyanite ions.

5. The composition of claim 1 wherein said oxidoreductase is selected from the group consisting of glucose oxidase, galactose oxidase, glycollate oxidase, lactate oxidase, L-gulonolactone oxidase, L-2-hydroxyacid oxidase, aldehyde oxidase, xanthine oxidase, D-aspartate oxidase, L-amino acid oxidase, D-amino acid oxidase, monoamine oxidase, pyridoxaminephosphate oxidase, diamine oxidase, and sulfite oxidase.

6. The composition of claim 1 wherein said oxidoreductase is glucose oxidase.

7. The composition of claim 1 wherein said substrates are specific to

the

particular oxidoreductase and are selected from D-glucose, D-

galactose, L-sorbose, ethanol, tyramine, 1, 4-diaminobutane, 6-hydroxy-L-nicotine, 6-hydroxy-D-nicotine, 2-aminophenol, glycollate, L-lactate, 2'-deoxy-D-Glucose, L-gulonolactone, L-galactonolactone, D-mannonolactone, L-2-hydroxyisocaproate, acetaldehyde, butyraldehyde, xanthine, D-aspartate, D-glutamate, L-amino acids and D-amino acids.

8. The composition of claim 1 wherein said oxidoreductase is glucose oxidase and said substrate is D-glucose.

9. The composition of claim 3 wherein the peroxidase enzyme is selected from lactoperoxidase, myeloperoxidase, salivary peroxidase, and chloroperoxidase.

10. The composition of claim 9 wherein the peroxidase is lactoperoxidase.

11. The composition of claim 1 wherein the catalase is derived from A. niger fermentation.

12. The composition of claim 1, wherein said composition comprises a fluid carrier, comprised of water, in an amount ranging from about 10%

to

about 90% by weight of the composition.

13. The composition of claim 1 further comprising a humectant selected from glycerine, propylene glycol, sorbitol (70% solution), polyethylene glycols, polypropylene glycols, and mixtures thereof.

14. The composition of claim 12 wherein said water comprises in the

range

from about 5% to about 50% by weight of said composition.

15. The composition of claim 1 further comprising a thickener selected from natural and synthetic water-soluble polymers selected from sodium carboxymethylcellulose, xanthan gum, carrageenan, locust bean gum, gum tragacanth, hydroxyethylcellulose, sodium alginate, starch, polyvinylpyrrolidone and polyacrylic acid and inorganic thickeners selected from magnesium aluminum silicate, hectorites and hydrated silicas.

16. The composition of claim 1 further comprising abrasives selected

from

the group consisting of calcium pyrophosphate, calcium carbonate, hydrated silica, aluminum hydroxide, dicalcium phosphate dihydrate, tricalcium phosphate, sodium metaphosphate, potassium metaphosphate, aluminum silicate, finely divided poly(methyl methacrylate), and mixtures thereof.

17. The composition of claim 1 wherein said composition further comprises

a physiologically acceptable buffer.

18. The composition of claim 17 wherein said physiologically acceptable buffer is selected from potassium phosphate, sodium phosphate, disodium phosphate, dipotassium phosphate, and mixtures thereof.

19. The composition of claim 1 further comprising additives selected from

the group comprising preservatives, whiteners, dyes, fluorides, antitartar and anticalculus agents, chlorophyll compounds, ammoniated materials, flavorings and sweeteners.

20. Method of making a dentifrice composition comprising the steps of: mixing together an oxidoreductase enzyme, an oxidoreductase enzyme substrate and a catalase, wherein said enzyme and substrate form

hydrogen

peroxide when reacted together, said hydrogen peroxide being formed at a rate of at least 100 micromoles per liter per minute said oxidoreductase enzyme being present in said composition in the amount of at least 1.0 Titrimetric Unit per gram of dentifrice and wherein said oxidoreductase enzyme and catalase are present in a fluid carrier in a ratio of about

50

Titrimetric Units oxidoreductase enzyme to 1.0 Baker Unit of catalase, to 1.0 Titrimetric Units oxidoreductase enzyme to 1.0 Baker Unit of catalase; and storing said mixture in an oxygen impervious container.

21. The method of claim 20 wherein the composition is made either under a

partial vacuum or in an oxygen free atmosphere after said oxidoreductase enzyme and an oxidoreductase enzyme substrate are added thereto.

22. The method claim 21 wherein said step of limiting the amount of oxygen in said composition is performed under an inert gas.

23. An anaerobically packaged composition with an antimicrobial system comprising: a fluid carrier comprising an oxidoreductase enzyme and an oxidoreductase enzyme substrate, wherein said enzyme and substrate form hydrogen peroxide when reacted together in the presence of oxygen, said hydrogen peroxide is formed at a rate of at least 100 micromoles per liter per minute; and catalase provided in sufficient amount to substantially reduce the amount of hydrogen peroxide in said composition;

said composition being packaged in an oxygen impervious package or container.

REP	US 2482724	Sep 1949	426010000	Baker
	US 2732988	Jan 1956	226051000	Feinstein
	US 2765233	Oct 1956	094178000	Sarett et al.
	US 3182432	May 1965	053112000	Canfield
	US 3430414	Mar 1969	053079000	Ludwig et al.
	US 3518809	Jul 1970	053112000	Ott
	US 4055931	Nov 1977	053022000	Myers
	US 4269822	May 1981	424050000	Pellico et al.
	US 4537764	Aug 1985	424050000	Pellico et al.
	US 4578265	Mar 1986	424050000	Pellico et al.
	US 4996062	Feb 1991	426008000	Lehtonen et al.
	US 5110609	May 1992	426402000	Lewis et al.
	US 5176899	Jan 1993	424050000	Montgomery
	DE 2520792	Nov 1976	426010000	
	GB 986178	Mar 1965	426404000	
NCL	NCLM: 424050000			
	NCLS: 053403000; 053405000; 053408000; 053432000; 424094400; 426404000; 426486000			
IC	ICM: A61K007-28			

ICS: A61K037-50  
EXF 053092000; 053400000; 053405000; 053408000; 42400000; 424094400;  
426404000; 426486000  
ARTU 125

L13 ANSWER 9 OF 13 WPIDS COPYRIGHT 2001 DERWENT INFORMATION LTD  
AN 1989-326272 [45] WPIDS  
DNC C1989-144419  
TI 5-C-hydroxy methyl-aldo hexose cpds. prepn. - by enzymatic oxidn. with  
e.g. **galactose oxidase**, followed by condensation with  
formaldehyde and base.

DC B03 D13 E13  
IN HILER, G D; KLUESENER, B W; MAZUR, A W; STIPP, G K  
PA (PROC) PROCTER & GAMBLE CO  
CYC 8

PI EP 341063 A 19891108 (198945)\* EN 14p  
DK 8902235 A 19891106 (199003)  
FI 8902144 A 19891106 (199006)  
AU 8933992 A 19891109 (199008)  
JP 02084190 A 19900326 (199018)  
US 5104797 A 19920414 (199218) 11p  
NZ 228994 A 19921125 (199305) C12P019-02  
FI 91069 B 19940131 (199408) C07H015-04  
EP 341063 B1 19940316 (199411) EN 20p C12P019-02  
DE 68913801 E 19940421 (199417) C12P019-02  
PH 26765 A 19920928 (199634) A23L001-09

ADT EP 341063 A EP 1989-304505 19890504; JP 02084190 A JP 1989-113999  
19890506; US 5104797 A US 1989-337725 19890417; NZ 228994 A NZ  
1989-228994

19890504; FI 91069 B FI 1989-2144 19890504; EP 341063 B1 EP 1989-304505  
19890504; DE 68913801 E DE 1989-613801 19890504, EP 1989-304505 19890504;  
PH 26765 A PH 1989-38609 19890504

FDT FI 91069 B Previous Publ. FI 8902144; DE 68913801 E Based on EP 341063  
PRAI US 1988-190485 19880505; US 1989-337725 19890417

REP 2.Jnl.Ref; A3...9036; EP 341062; No-SR.Pub

IC ICM A23L001-09; C07H015-04; C12P019-02

ICS A23L001-23; A23L001-236; C07H001-00; C07H003-10; C07H019-01;  
C07N001-00; C12N009-04; C12P019-44

AB EP 341063 A UPAB: 19930923

Prepn. comprises (a) reacting under agitation an aq. soln. pref. having  
pH6-8 and at 1-50 deg.C, comprising (i) 1-50% of at least 1 D-aldohexose  
based cpd. and (ii) 1,000-1,000,000 unit activity of the enzyme  
D-aldohexose:oxygen 6-oxidoreductase per mole of D-aldbhexase based cpd.,  
(b) reacting the obtd. soln. with 1-40 mol equivalents of formaldehyde

and  
1-13 mol equivalents of at least 1 base of NaOH, Ca(OH)2 and KOH, at  
15-40

deg.C and pH12-13 and (c) purifying the obtd. aq. 5-C-hydroxymethyl  
-D-aldohexase based cpd. contg. soln., pref. by dewatering and  
crystallising the cpd. The condensn. prod. of step (c) is additionally  
(d) hydrolysed with 1-10 molar equivalents of at least 1 of H2SO4, HNO3,  
HCl, perchloric acid, phosphoric acid, methanesulphonic acid and  
trifluoromethane sulphonic acid while maintaining the reaction mixt. at

20  
deg.C to boiling, pref. 80-100 deg.C and (e) residual acid is removed

from  
the reaction mixt, pref. by neutralisation and then pptn.

USE - Derivs. of 5-C-hydroxymethyl-D-hexose cpds. are used as  
replacement for sugar, esp. in **baked** goods.

0/0

FS CPI

FA AB; DCN

MC CPI: B07-A02; D03-H01A; E07-A02H

L13 ANSWER 10 OF 13 IFIPAT COPYRIGHT 2001 IFI  
AN 1536343 IFIPAT;IFIUDB;IFICDB

TI PREPARATION OF ENZYMATICALLY ACTIVE FORMULATION EMBEDDED IN SILICA  
GEL

INF Klefenz, Heinrich, Hochdorf-Assenheim, DE

Sanner, Axel, Frankenthal, DE

Tschang, Chung-Ji, Frankenthal, DE

Zahn, Wolfgang, Altrip, DE

IN KLEFENZ HEINRICH (DE); SANNER AXEL (DE); TSCHANG CHUNG-JI (DE); ZAHN  
WOLFGANG (DE)

PAF BASF Aktiengesellschaft, DE

PA BASF AG DE (7016)

EXNAM Lovering, Richard D

AG Keil & Weinkauff

PI US 4461832 19840724 (CITED IN 007 LATER PATENTS)

AI US 1982-376597 19820510

XPD 24 Jul 2001

RLI US 1980-125035 19800227 CONTINUATION ABANDONED

PRAI DE 1979-2911776 19790326

FI US 4461832 19840724

DT UTILITY; EXPIRED

FS CHEMICAL

MRN 004226 MFN: 0596

AB A process for the preparation of an enzymatically active formulation embedded in silica gel, wherein an aqueous mixture of an enzymatically active formulation and a dissolved alkali metal silicate and/or ammonium silicate is suspended in an organic, water-immiscible fluid and then converted to a water-insoluble gel.

CLMN 11

ECLM 1. A PROCESS FOR THE PREPARATION OF AN ENZYMATICALLY ACTIVE FORMULATION EMBEDDED IN SILICA GEL WHICH COMPRISES: SUSPENDING AN AQUEOUS MIXTURE CONSISTING OF AN ENZYMATICALLY ACTIVE FORMULATION AND A DISSOLVED ALKALI METAL SILICATE AND/OR AMMONIUM SILICATE AS DROPLETS IN A STIRRED

ORGANIC,

WATER-IMMISCIBLE FLUID AND THEN CONVERTING SAID SILICATE TO A WATER-INSOLUBLE GEL.

ACLM 2. A process as claimed in claim 1, wherein the enzymatically active formulation used consists of active cells or cell fragments of microbiological, vegetable, animal or human origin.

3. A process set forth in claim 1, wherein the formation of a suspension is assisted by using a suspending agent.

4. A process as set forth in claim 1, 2 or 3, wherein the gelling is effected by lowering the pH by means of an agent which is soluble in water and in the organic water-immiscible fluid.

5. A process of claim 4 wherein said agent is an organic acid.

6. A process as set forth in claim 1, wherein the gelling is effected in the presence of an inert material.

7. A process as claimed in claim 1, wherein said enzymatically active formulation embody as micro-organisms one of Streptomyces, Arthrobacter or Bacillus microorganisms, or Escherichia coli, Saccharomuces, Curvularia lunata, or Aspergillus ochraceus.

8. A process as claimed in claim 1, wherein the enzymatically active formulation embody of one of trypsin, chymotrypsin, pancreatin, Alpha - and Beta -amylase, ribonucleases, desoxyribonucleases, cellulase, maltase, pectinase, chitinase, pepsin, bromelain, keratinase, amyloglycosidase, lipase, cholinesterase, lecithinase, phosphatase, alginase, asparaginase, glutaminase, urease, lactase,

penicillin-amidase,

penicillinase, glucose-isomerase, glucose-oxidase, catalase, peroxidase, lipoxidase, xanthin-oxidase, cytochrome-reductase, lactic acid oxidase, aminoacid oxidase, rennin, ficin, subtilisin, tannase, phenol-oxidase, pullulanase, isoamylase, hexokinase, galactose-oxidase

, diaphorase, aldolase, glycollic acid oxidase, luciferase,

aldehyde-oxidase, naringinase, uricase, glutathione-reductase,

nitrito-reductase, nitrate-reductase, succinic acid dehydrogenase,

catechol-oxidase, Beta -fructosidase, aminoacid acylase and urokinase.

9. A process as claimed in claim 1, wherein said enzymatically active formulation comprises baker's yeast.

10. A process as claimed in claim 1, wherein the embedded enzymatic formulation is immobilized by the embedding thereof in said gel.  
11. A process for the preparation of an enzymatically active formulation embedded in silica gel which comprises: suspending an aqueous mixture consisting of an enzymatically active formulation, soluble sulfates or phosphates and a dissolved alkali metal silicate and/or ammonium

silicate

as droplets in a stirred organic, water-immiscible fluid, then

converting

said silicate to a water-insoluble gel, and thereafter treating the product obtained with a solution of salt of which the cation forms a sparingly soluble sulfate or phosphate.

REP	US 2982749	May 1961	526910000X	Friedrich et al.
	US 3069370	Dec 1962	264004100X	Jensen et al.
	US 3791987	Feb 1974	264004000X	Fanger
	US 3948866	Apr 1976	252009000X	Pennewiss et al.
	US 3954678	May 1976	252062530X	Marquisee
	US 4011096	Mar 1977	106288000B	Sandell
	US 4164613	Aug 1979	526201000	Hoene et al.
	DE 2625704	Dec 1976		
	GB 1267685	Mar 1972	435176000	

REN Rose et al.: The Condensed Chemical Dictionary, 4th Edition, Reinhold Publ. Corp., 1950, p. 710.

NCL NCLM: 435176000

NCLS: 264004100; 264004300; 424094300; 424094400; 424094500; 424094600;  
424094610; 424094630; 424094660; 427213300; 427213320; 435182000;  
436527000; 436535000; 436823000; 436826000

IC ICM: C12N011-14

ICS: B01J013-02

EXF 264004100; 264004300; 427213300; 427213320; 435176000

ARTU 223

L13 ANSWER 11 OF 13 GENBANK.RTM. COPYRIGHT 2001

LOCUS (LOC): SCE20 GenBank (R)

GenBank ACC. NO. (GBN): AL136058

CAS REGISTRY NO. (RN): 252838-53-2

SEQUENCE LENGTH (SQL): 33820

MOLECULE TYPE (CI): DNA; linear

DIVISION CODE (CI): Bacteria

DATE (DATE): 7 Jan 2000

DEFINITION (DEF): Streptomyces coelicolor cosmid E20.

KEYWORDS (ST): 2-hydroxyhepta-2,4-diene-1,7-dioate isomerase; amino acid AB; transporter protein integral, membrane component; amino acid AB; transporter protein, ATP-binding component; arr; chb; glycosyl; transferase; GntR-family transcriptional regulatory protein; helicase; IclR-family transcriptional regulator; lipoprotein; LysR-family transcriptional regulatory protein; membrane protein; N-acetylglucosamine-6-phosphate deacetylase; oxidoreductase; rifampin ADP-ribosyl transferase; secreted chitin binding protein; secreted endoglucanase; secreted protein; transcriptional regulatory protein; transmembrane efflux protein; tRNA Lys

SOURCE: Streptomyces coelicolor A3(2).

ORGANISM (ORGN): Streptomyces coelicolor A3(2)

Bacteria; Firmicutes; Actinobacteria;

Actinobacteridae;

Actinomycetales; Streptomycineae; Streptomycetaceae;  
Streptomyces

NUCLEIC ACID COUNT (NA): 4825 a 12217 c 12007 g 4771 t

COMMENT:

Notes:

Streptomyces coelicolor sequencing at The Sanger Centre is funded by the BBSRC and Beowulf Genomics



Details of *S. coelicolor* sequencing at the Sanger Centre are available on the World Wide Web.  
 (URL; [http://www.sanger.ac.uk/Projects/S\\_coelicolor/](http://www.sanger.ac.uk/Projects/S_coelicolor/))  
 CDS are numbered using the following system eg SC7B7.01c. SC (*S. coelicolor*), 7B7 (cosmid name), .01 (first CDS), c (complementary strand).

The more significant matches with motifs in the PROSITE database are also included but some of these may be fortuitous.  
 The length in codons is given for each CDS.

Usually the highest scoring match found by fasta -o is given for CDS which show significant similarity to other CDS in the database.  
 The position of possible ribosome binding site sequences are given where these have been used to deduce the initiation codon.

Gene prediction is based on positional base preference in codons using a specially developed Hidden Markov Model (Krogh et al., Nucleic Acids Research, 22(22):4768-4778(1994)) and the FramePlot program of Bibb et al., Gene 30:157-66(1984) as implemented at <http://www.nih.go.jp/jun/cgi-bin/frameplot.pl>.

CAUTION: We may not have predicted the correct initiation codon. Where possible we choose an initiation codon (atg, gtg, ttg or (att)) which is preceded by an upstream ribosome binding site sequence (optimally 5-13bp before the initiation codon). If this cannot be identified we choose the most upstream initiation codon.

IMPORTANT: This sequence MAY NOT be the entire insert of the sequenced clone. It may be shorter because we only sequence overlapping sections once, or longer, because we arrange for a small overlap between neighbouring submissions.

Cosmid E20 Overlaps with cosmid E6 on the AseI-E genomic restriction fragment.

REFERENCE: 1 (bases 1 to 33820)  
 AUTHOR (AU): Redenbach,M.; Kieser,H.M.; Denapate,D.; Eichner,A.; Cullum,J.; Kinashi,H.; Hopwood,D.A.  
 TITLE (TI): A set of ordered cosmids and a detailed genetic and physical map for the 8 Mb *Streptomyces coelicolor*

A3(2)  
 chromosome  
 JOURNAL (SO): Mol. Microbiol., 21 (1), 77-96 (1996)  
 OTHER SOURCE (OS): CA 125:159753

REFERENCE: 2 (bases 1 to 33820)  
 AUTHOR (AU): Seeger,K.J.; Harris,D.  
 JOURNAL (SO): Unpublished

REFERENCE: 3 (bases 1 to 33820)  
 AUTHOR (AU): Thomson,N.R.; Parkhill,J.; Barrell,B.G.; Rajandream,M.A.  
 TITLE (TI): Direct Submission  
 JOURNAL (SO): Submitted (07-JAN-2000) *Streptomyces coelicolor* sequencing project, Sanger Centre, Wellcome Trust Genome Campus, Hinxton, Cambridge CB10 1SA E-mail: barrell@sanger.ac.uk Cosmids supplied by Prof. David

A.  
 Hopwood, [3] John Innes Centre, Norwich Research Park, Colney, Norwich, Norfolk NR4 7UH, UK

#### FEATURES (FEAT):

Feature Key	Location	Qualifier
source	1..33820	/organism="Streptomyces coelicolor A3(2)" /strain="A3(2)" /db-xref="taxon:100226" /clone="cosmid E20"
gene	1..107	/gene="SCE20.01c"
CDS	<1..107	/gene="SCE20.01c" /note="SCE20.01c, unknown, partial CDS, len: 32 aa."

gene 118..1047  
CDS 118..1047

/codon-start=3  
/transl-table=1  
/product="hypothetical protein  
SCE20.01c."  
/protein-id="CAB65557.1"  
/db-xref="GI:6689160"  
/translation="SAMP SMAVPRGLAVKGAGPM  
YRTEKDRHNLPAFG"  
/gene="SCE20.02"  
/gene="SCE20.02"  
/note="SCE20.02, probable amino  
acid ABC transporter protein,  
solute-binding component, len: 309  
aa. Similar to several including:  
Salmonella typhimurium  
SW:ARGT-SALTY(EMBL:V01368)  
lysine-arginine-ornithine-binding  
periplasmic protein precursor (260  
aa), fasta scores opt: 251  
z-score: 294.5 E(): 5.1e-09 25.8%  
identity in 279 aa overlap and  
Rhizobium sp. (strain NGR234)  
SW:Y4TE-RHISN(EMBL:AE000098)  
probable amino-acid ABC  
transporter periplasmic binding  
protein Y4TE (300 aa), fasta  
scores opt: 599 z-score: 691.7  
E(): 3.8e-31 35.4% identity in 297  
aa overlap. Contains a Pfam match  
to entry PF00497 SBP-bac-3,  
Bacterial extracellular  
solute-binding proteins, family 3.  
Also contains an N-terminal  
possible membrane spanning  
hydrophobic domain."

misc-feature 184..993

/codon-start=1  
/transl-table=11  
/product="probable amino acid ABC  
transporter protein,  
solute-binding component."  
/protein-id="CAB65558.1"  
/db-xref="GI:6689161"  
/translation="MAPPHRTPPWGISDQRPRTA  
GPTRRSLLAGVAALGALGAAGCSR  
VATASDVKGGDLLDRLRAQQGVARLGIAGEVPFGY  
IDKNGELTGEAPELAKVIFKRLGV  
DRVQVPVTEFGSLIPGLASQQFDVVAAGMYINPD  
RCQQVIFSDPDYQMLDAYIVRKGN  
PLGLHNYRDVVKKAKFATGTGYAEIAYAVEHG  
Y KEDDILIVPDQVAGLNAVESGRVD  
VFAGTALTVRDVVKSSKAEATEAFAPLVDGKPH  
VDGGGFAFRPDETNLRFNVELQ  
KLKKSCELLRLKPFQFTQNEMTDLTAKELCGG"  
/gene="SCE20.02"  
/note="Pfam match to entry PF00497  
SBP-bac-3, Bacterial extracellular  
solute-binding proteins, family 3,  
score 77.30, E-value 3.2e-19"

gene 1044..1745  
CDS 1044..1745

/gene="SCE20.03"  
/gene="SCE20.03"  
/note="SCE20.03, amino acid ABC  
transporter protein, integral  
membrane component, len: 233 aa.  
Highly similar to many including:  
Escherichia coli  
SW:GLNP-ECOLI(EMBL:X14180)

		glutamine transport system permease protein GlnP (219 aa), fasta scores opt: 377 z-score: 445.7 E(): 1.9e-17 27.8% identity in 212 aa overlap and Rhizobium sp. (strain NGR234) SW:Y4TF-RHISN(EMBL:AE000098) probable amino-acid ABC transporter permease protein Y4TF (238 aa), fasta scores opt: 570 z-score: 667.5 E(): 8.5e-30 45.1% identity in 204 aa overlap. Contains a Pfam match to entry PF00528 BPD-transp, Binding-protein-dependent transport systems inner membrane component. Also contains a possible N-terminal signal sequence and possible membrane spanning hydrophobic domains." /codon-start=1 /transl-table=11 /product="probable amino acid ABC transporter protein, integral membrane component." /protein-id="CAB65559.1" /db-xref="GI:6689162" /translation="MTSGLWELVLQGVWVTVQLL FFSSLLATAVSFVVGIARSHRLWI VRFLAGFYTEVFRGTSALVMIFWVFFVLPPAFGW QLVPMWAGTLALGLTYGAYGSEIV RGS�AAVDPAQKEGGIALSFTPWQRMKLILLPQA VPEMIPPFSNLLIELLKGTALVSI MGMGDLAFSANLVRLLALQESAIEITYVLLIYFVI AFLLTRVMRGLEKKLKAGVGKAPK KKTAAVRVPEGSGVS" /gene="SCE20.03" /note="Pfam match to entry PF00528 BPD-transp, Binding-protein-dependent transport systems inner membrane component, score 44.30, E-value 2.7e-09"
misc-feature	1356..1586	
gene	1742..2392	/gene="SCE20.04"
CDS	1742..2392	/gene="SCE20.04" /note="SCE20.04, probable amino acid ABC transporter protein, integral membrane component, len: 216 aa. Highly similar to several ABC transporter permeases including: Escherichia coli SW:GLNP-ECOLI(EMBL:X14180) glutamine transport system permease protein GlnP (219 aa), fasta scores opt: 378 z-score: 454.8 E(): 6e-18 33.2% identity in 217 aa overlap and Rhizobium sp. (strain NGR234) SW:Y4TG-RHISN(EMBL:AE000098) probable amino-acid ABC transporter permease protein Y4TG (231 aa), fasta scores opt: 667 z-score: 792.8 E(): 0 47.6% identity in 208 aa overlap. Contains multiple possible membrane spanning hydrophobic

domains and a match to entry  
PF00528 BPD-transp,  
Binding-protein-dependent  
transport systems inner membrane  
component."  
/codon-start=1  
/transl-table=11  
/product="probable amino acid ABC  
transporter protein, integral  
membrane component."  
/protein-id="CAB65560.1"  
/db-xref="GI:6689163"  
/translation="MKWDWSAVSDFMPHFWDGGL  
VTLQILVLGSLVSFGLGLVWALLM  
RVPSRWVTWPVGVTVEFVRNTPLLVLFFLYVL  
PEWNITFSALTGTGVVAIGLHYSTY  
TMQVYRAGIEGVPVGQWEAATALNLPMTWTAV  
ILPQAIRRVTPALGNVVISMLKDT  
PLLMAITVLEMLGEARLFSQQNFQFTEPLTVIGV  
AFIVISYLAALRALERRLAH"  
/misc-feature 2069..2290  
/gene="SCE20.04"  
/note="Pfam match to entry PF00528  
BPD-transp,  
Binding-protein-dependent  
transport systems inner membrane  
component, score 37.50, E-value  
3.1e-07"  
/gene="SCE20.05"  
/gene="SCE20.05"  
/note="SCE20.05, probable amino  
acid ABC transporter protein,  
ATP-binding component, len: 261  
aa. Highly similar to many  
ATP-binding transport protein  
including: Bacillus  
stearothermophilus  
SW:GLNQ-BACST(EMBL:M61017)  
glutamine transport ATP-binding  
protein GLNQ (242 aa), fasta  
scores opt: 758 z-score: 851.7  
E():0 47.5% identity in 242 aa  
overlap and Thermotoga maritima  
TR:Q9WZ60(EMBL:AE001734) amino  
acid ABC transporter, ATP-binding  
protein (242 aa), fasta scores  
opt: 829 z-score: 930.4 E(): 0  
54.5% identity in 242 aa overlap.  
Also similar to several other  
Streptomyces coelicolor transport  
proteins e.g.  
TR:O50495(EMBL:AL020958) glutamate  
uptake system ATP-binding protein,  
GluA (258 aa), fasta scores opt:  
775 z-score: 683.3 E(): 1.1e-32  
48.2% identity in 251 aa overlap.  
Contains Prosite hits to PS00211  
ABC transporters family signature  
and PS00017 ATP/GTP-binding site  
motif A (P-loop). Also contains a  
Pfam match to entry PF00005  
ABC-tran, ABC transporter."  
/codon-start=1  
/transl-table=11  
/product="probable amino acid ABC  
transporter protein, ATP-binding  
component."

misc-feature 2538..2561

misc-feature 2580..3080

misc-feature 2850..2894

gene 3250..4107  
CDS 3250..4107

misc-feature 3550..4074

```
/protein-id="CAB65561.1"
/db-xref="GI:6689164"
/translation="MPTDTLPNPEKSPERSSGEL
IRLEQVTKRFGDNTVLDHLDLDFSV
AGKHVTLIGPSGSGKTTILRLMTLLKPDEGTIT
VDGQKLFPAEKERREVRKQIGMV
FQQFNLFNMSVLRNITEAPVTVLGMPKDEAVER
AKGLLDVGLADKCDARPAQLSGG
QQQVVAIARALAMRPKVLLDEVTSALDPELVAG
VLDLLRDIARSTDITMLCVTHEMN
FARDISDQVLMFDSGRVIEAGAPEKIFSEPEHDR
TREFLSAVL"
/gene="SCE20.05"
/note="PS00017 ATP/GTP-binding
site motif A (P-loop)"
/gene="SCE20.05"
/note="Pfam match to entry PF00005
ABC-tran, ABC transporter, score
149.60, E-value 5.4e-41"
/gene="SCE20.05"
/note="PS00211 ABC transporters
family signature"
/gene="SCE20.06"
/gene="SCE20.06"
/note="SCE20.06, possible
IclR-family transcriptional
regulator, len: 285 aa, Almost
identical to a gene fragment
which, like SCE20.06, is located
upstream of the Streptomyces
lividans lysT tRNA gene:
TR:Q54411(EMBL:X52073) (200 aa),
fasta scores opt: 1250 z-score:
1464.9 E():0 99.5% identity in 200
aa overlap. Also similar to
Streptomyces coelicolor
TR:Q9X9U3(EMBL:AL096823) putative
transcriptional regulator (241
aa), fasta scores opt: 597
z-score: 704.2 E(): 7.7e-32 43.2%
identity in 236 aa overlap.
Contains a Pfam match to entry
PF01614 IclR, Bacterial
transcriptional regulator with a
putative helix-turn-helix motif
situated between residues 41..62
(+3.07 SD)."
/codon-start=1
/transl-table=11
/product="putative IclR-family
transcriptional regulator."
/protein-id="CAB65562.1"
/db-xref="GI:6689165"
/translation="MRAPKSAQHPLPGSLLAAIV
NAGMHQRAQGETTVALQHKPTAPH
HSTEDALRVLETVARHTSGVTDTEIARES
GIGTERLTLLRMLRREAYVEQTADGAYV
TGEALARLGSAGGREQALREKLQRTLEGL
RDSVGAAYVISRYVDGEVSVTQYADSPAA
PRVNEWVDFRVS AHATAVGKSLLTQLDH
AGRRDLARHRMARLTSRTITSDKLLLSRL
ESQPATVPVLDLQEYAVGTVCAAVPITAG
SAVGC LALSLPVEHAHRLRRAADELNRSA
APVLLSLAI"
/gene="SCE20.06"
/note="Pfam match to entry PF01614
```

tRNA  
gene  
CDS

4197..4270  
complement(4316..4921)  
complement(4316..4921)

IclR, Bacterial transcriptional  
regulator, \score=21.90, E-value  
6e-08"

/note="tRNA Lys anticodon CTT"  
/gene="chb"  
/gene="chb"  
/note="SCE20.07c, chb, secreted  
chitin binding protein, len: 201  
aa. Highly similar to several  
including: Streptomyces  
olivaceoviridis (Streptomyces  
corchorusii)  
TR:Q54501(EMBL:X78535) chitin  
binding protein precursor Chb1  
(201 aa), fasta scores opt: 1185  
z-score: 1303.0 E():0 83.7%  
identity in 202 aa overlap and  
Streptomyces reticuli  
TR:O87962(EMBL:Y14315) chitin  
binding protein (Chb2) (201 aa),  
fasta scores opt: 1161 z-score:  
1276.8 E(): 0 79.1% identity in  
201 aa overlap. Contains a  
possible N-terminal signal  
sequence."

/codon-start=1  
/transl-table=11  
/product="secreted chitin binding  
protein."  
/protein-id="CAB65563.1"  
/db-xref="GI:6689166"  
/translation="MRTRTKLYAAALGMATTGAL  
VLSSGGASGHGYTDLPVSRQKVCQ  
NGTVGGCGAIQWEPQSVEGPKGFASGPADGTIC  
SAGHGSFAALDSPKQPNGQAWPTT  
RVNGGQSYTFRWQFTARHATTDKYYVTKPGWNQ  
NHNLARS DLNLT PFFTVPYGGKQP  
PATLSHSGTLP SGLSGHHVILAVVTVHDTGNAFY  
ACSDVTF"

gene  
CDS

complement(5042..6193)  
complement(5042..6193)

/gene="SCE20.08c"  
/gene="SCE20.08c"  
/note="SCE20.08c, possible  
membrane protein, len: 383 aa.  
Contains possible membrane  
spanning hydrophobic domains."  
/codon-start=1  
/transl-table=11  
/product="putative membrane  
protein"  
/protein-id="CAB65564.1"  
/db-xref="GI:6689167"  
/translation="MSTTSSTNPETAPAAPEESA  
AAAGQRSARLIHNEATTEIPVHLL  
FRDDPD PAPVPLRPVAVARRPGPGERTGARPGAR  
RPVAARPRPAPQVDPELTERPGRV  
LPGAAGVAAGLCGAAGCAATSWWAGLVPPLAAQA  
LGLPAYAGAGLGPQWAAAYAAAGA  
LGMFGFGLARGRTGRAWVLGLFGRYRGTVRRTG  
LMWVNPLLLRRRVDVRLRHWRSEP  
MPAADGNGVALRAVTLVVWRVRDTAKATLGVEDH  
ETYLRECVAAALARPVEPLGTVR  
SSADVAGDTLTRLVAADAAAPVGLEVFSVRPVRVE  
YAPEVAAAMHRRRIAALDAAQRAS  
VLT SVVDSVEDTVTRLTMRGLVELDDYERKVLVK  
DLTVAFCAGRGDTGH"

gene

complement(6250..7311)

/gene="SCE20.09c"

CDS

comp[REDACTED]nt (6250..7311)

/gene="SCE20.0"  
/note="SCE20.0", possible  
membrane protein, len: 353 aa.  
Contains a possible membrane  
spanning hydrophobic domain and is  
rich in the amino acid Ala."  
/codon-start=1  
/transl-table=11  
/product="putative membrane  
protein."  
/protein-id="CAB65565.1"  
/db-xref="GI:6689168"  
/translation="METPVFEEIEPASDCDCAGC  
RHWRRVLPAPAGRALAHFAAYRT  
LVAAAAAASATAVSVLGAGGAAALPAAHAHAHRP  
GVPAGDEPGTPQGPRGPLHGPVGR  
PAAPPAGAEIAGITRTEIIDRAKSWVAAKVPYSL  
TAYWSDGYRQDCSGFVSMAWKLPA  
NEWTGSLGTVADRITKEELQPGDILLFHNESDPQ  
KGSHVVFIFGG

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u

=> d 12

L13 ANSWER 12 OF 13 FROSTI COPYRIGHT 2001 LFRA  
AN 489211 FROSTI  
TI A composition comprising an enzyme having **galactose  
oxidase** activity and use thereof.  
IN Rouau X.; Schroder M.; Soe J.B.  
PA Danisco A/S  
SO PCT Patent Application  
PI WO 9903351 A1  
AI 19980716  
PRAI Denmark 19970718  
United States 19970722  
DT Patent  
LA English  
SL English

=> d 12 ab,bib

L13 ANSWER 12 OF 13 FROSTI COPYRIGHT 2001 LFRA  
AB A composition for **dough** and bread improvement is disclosed. It  
incorporates an enzyme with **galactose oxidase**  
activity and an oxidizable substrate for this enzyme. It is said to  
improve the rheological properties of flour **doughs** and the  
quality characteristics of bread products. Desirable quality  
characteristics include soft crumb structure, high specific volume, and  
freedom from staling within the expected shelf-life of fresh bread.  
**Galactose oxidase** acts as an oxidoreductase, and its  
use overcomes problems associated with use of cellulases or  
**hemicellulases** in flour **doughs**. Because the natural  
**galactose** content of cereal flours is very low, it is beneficial  
to include an oxidizable substrate in the formulation.  
AN 489211 FROSTI  
TI A composition comprising an enzyme having **galactose  
oxidase** activity and use thereof.  
IN Rouau X.; Schroder M.; Soe J.B.  
PA Danisco A/S  
SO PCT Patent Application  
PI WO 9903351 A1  
AI 19980716  
PRAI Denmark 19970718

DT United States 70722  
LA Patent  
SL English  
SL English

=> d 13 ab,bib

L13 ANSWER 13 OF 13 FROSTI COPYRIGHT 2001 LFRA  
AB A composition for **dough** and bread improvement is disclosed. It incorporates an enzyme with **galactose oxidase** activity and an oxidizable substrate for this enzyme. It is said to improve the rheological properties of flour **doughs** and the quality characteristics of bread products. Desirable quality characteristics include soft crumb structure, high specific volume, and freedom from staling within the expected shelf-life of fresh bread. **Galactose oxidase** acts as an oxidoreductase, and its use overcomes problems associated with use of cellulases or **hemicellulases** in flour **doughs**. Because the natural **galactose** content of cereal flours is very low, it is beneficial to include an oxidizable substrate in the formulation.

AN 526332 FROSTI  
TI A composition comprising an enzyme having **galactose oxidase** activity and use thereof.

IN Rouau X.; Schroder M.; Soe J.B.  
PA Danisco A/S  
SO European Patent Application  
PI EP 999752 A1  
WO 9903351 19990128  
AI 19980716  
PRAI Denmark 19970718  
United States 19970722

DT Patent  
LA English  
SL English